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(54) Title: PROSTATE CANCER–ASSOCIATED GENES

(57) Abstract

The invention provides novel prostate cancer–associated genes and polypeptides encoded by those genes. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating or preventing diseases.

PROSTATE CANCER-ASSOCIATED GENES

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10   **TECHNICAL FIELD**

The invention relates to a method for analyzing gene expression patterns. The

invention also relates to eight prostate cancer-associated genes identified by the method

and their corresponding polypeptides and to the use of these biomolecules in diagnosis,

15 prognosis, treatment, prevention, and evaluation of therapies for diseases, particularly

diseases associated with cell proliferation, such as cancer.

BACKGROUND OF THE INVENTION

20   The DNA sequences of many human genes have been determined, but for many of

these genes, their biological function, and in particular their relationship to disease, is

unknown or poorly understood. Current laboratory and computational methods to

determine or predict the possible functions of newly-sequenced genes are slow and

expensive. Thus, new methods that provide additional information on function are

25 desirable.

Prostate cancer is a common malignancy in men over the age of 50, and the

incidence increases with age. In the US, there are approximately 132,000 newly

diagnosed cases of prostate cancer and more than 33,000 deaths from prostate cancer each

year. The occurrences of prostate cancer vary among different regions in the world. For

30 example, there are 14 deaths per 100,000 men per year in the US, compared with 22 in

Sweden and 2 in Japan.

Genes known to be involved in prostate cancer, such as prostate-specific antigen

(PSA), prostatic acid phosphatase (PAP), kallikrein, seminal plasma protein, and prostate-specific tranglutaminase, have been used or proposed as the basis for diagnostic and prognostic tests as well as therapeutic targets. In particular, prostate-specific antigen (PSA) is a protease used in diagnosis for prostate cancer (Morris, D. L. et al. (1998) J.

5   Clin. Lab. Anal. 12: 65-74). Prostatic acid phosphatase (PAP) is a phosphomonoesterase synthesized in the prostate and secreted into the seminal plasma under androgenic control (Ostrowski, W. S. and R. Kuciel (1994) Clin. Chim. Acta 226:121-129), and has been used in diagnostic tests for prostate cancer and in prognostic tests for metastatic cancer (Presti, J. C., Jr. and P. R. Carroll (1996) Semin Urol Oncol 14(3): 134-138). Kallikrein is a

10   protease expressed specifically in the prostate and has 80% sequence similarity with PSA (Corey, E., K. R. et al. (1997) Urology 50: 567-572). Kallikrein is being evaluated for use in diagnostic tests for prostate cancer (Pannek, J. and Partin, A. W. (1997) Oncology 11: 1273-1282). Seminal plasma protein is a prostate-specific secreted protein with activity similar to inhibin, a member of the transforming growth factor superfamily implicated in

15   prostate cancer (Mbikay, M., S. et al. (1987) DNA 6: 23-29; Thomas, T. Z. et al. (1998) Prostate 34: 34-43); deletion of the inhibin alpha gene in male rats results in development of primary gonadal granulosa/Sertoli cell tumors (Mellor, S. L.et al. (1998) J. Clin. Endocrinol. Metab. 83: 969-975). Prostate-specific transglutaminase catalyzes post-translational protein cross-linking, and exhibits differential expression in prostate

20   cancer cell lines (Dubbink, H. J. (1996) Biochem. J. 315: 901-908).

The diagnostic sensitivity and specificity and the prognostic accuracy of the tests based on the known genes are substantially less than 100 percent. For example, about 20 percent of the patients undergoing prostatectomy for prostate cancer have normal levels of PSA (Presti and Carroll, supra). Therefore, identification of novel genes and polypeptides

25   that are markers of and potential therapeutic targets for prostate cancer is desirable.

The present invention satisfies a need in the art by providing new compositions which are useful in diagnosis, prognosis, treatment, prevention, and evaluation of therapies for diseases, particularly diseases associated with cell proliferation, such as cancer. We have implemented a method for analyzing gene expression patterns and have identified

30   eight human prostate cancer-associated genes by their coexpression with known prostate cancer-specific genes.

SUMMARY OF THE INVENTION

In one aspect, the present invention provides a method for identifying biomolecules, such as polynucleotides or polypeptides, useful in the diagnosis, prognosis, treatment, prevention, and evaluation of therapies for diseases, particularly diseases associated with cell proliferation such as cancer, more particularly prostate cancer. The method can also be employed for elucidating genes involved in a common regulatory pathway.

The method comprises first characterizing expression patterns of polynucleotides that are expressed in a plurality of cDNA libraries. The expressed polynucleotides comprise genes of known and unknown functions. Second, the expression patterns of one or more function-specific genes are compared with the expression patterns of one or more of the genes of unknown function to identify a subset of novel genes which have similar expression patterns to those of the function-specific genes.

The method compares the expression pattern of two genes by first generating an occurrence vector for each gene. The vector comprises entries for each gene wherein a gene's presence in a cDNA library is represented by a one and a gene's absence by a zero. The vectors are then analyzed to determine whether the expression patterns of any of the genes are similar. Expression patterns are similar if a particular coexpression probability threshold is met. Preferably, the coexpression probability threshold is less than 0.001, and more preferably less than 0.00001.

In a preferred embodiment, the function-specific genes are prostate cancer-specific gene sequences including prostate-specific antigen (PSA), prostatic acid phosphatase (PAP), kallikrein, seminal plasma protein, prostate-specific tranglutaminase, and the like. These prostate cancer-specific genes are used to identify other polynucleotides of unidentified function that are predominantly coexpressed with the prostate cancer-specific genes. The polynucleotides analyzed by the present invention can be expressed sequence tags (ESTs), assembled sequences, full length gene coding sequences, introns, regulatory regions, 5' untranslated regions, 3' untranslated regions and the like.

In a second aspect, the invention entails a substantially purified polynucleotide identified by the method of the present invention as being associated with prostate cancer. In particular, the polynucleotide comprises a sequence selected from the group consisting

of SEQ ID NOs: 1-8 or its complement or a variant having at least 70% sequence identity to SEQ ID NOs: 1-8 or a polynucleotide that hybridizes under stringent conditions to SEQ ID NOs: 1-8 or a polynucleotide encoding SEQ ID NOs: 9 and 10. The present invention also entails a polynucleotide comprising at least 18 consecutive nucleotides of a sequence

5   provided above. The polynucleotide is suitable for use in diagnosis, treatment, prognosis, or prevention of a cancer, and in particular, prostate cancer. The polynucleotide is also suitable for the evaluation of therapies for cancer.

In another aspect, the invention provides an expression vector comprising a polynucleotide described above, a host cell comprising the expression vector, and a

10   method for detecting a target polynucleotide in a sample.

In a further aspect, the invention provides a substantially purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:9 and SEQ ID NO:10. The invention also provides a substantially purified polypeptide having at least 85% identity to SEQ ID NOs:9-10. Additionally, the invention also

15   provides a sequence with at least 6 sequential amino acids of SEQ ID NOs:9-10.

The invention also provides a method for producing a substantially purified polypeptide comprising the amino acid sequence referred to above, and antibodies, agonists, and antagonists which specifically bind to the polypeptide. Pharmaceutical compositions comprising the polynucleotides or polypeptides of the invention are also

20   contemplated. Methods for producing a polypeptide of the invention and methods for detecting a target polynucleotide complementary to a polynucleotide of the invention are also included.

In a general aspect, the invention entails a method for identifying biomolecules useful in the diagnosis or treatment of a disease or condition. The method comprises a)

25   examining expression patterns of a plurality of biomolecules that are expressed in a plurality of cDNA libraries, said expressed biomolecules comprising one or more disease-specific biomolecules and one or more biomolecules of unknown function; and b) comparing the expression patterns of said disease-specific biomolecules with the expression patterns of the biomolecules of unknown function to identify a subset of the

30   biomolecules of unknown function which have similar expression patterns to those of disease-specific biomolecules.

BRIEF DESCRIPTION OF THE SEQUENCE LISTING

The Sequence Listing provides exemplary prostate cancer-associated sequences including polynucleotide sequences, SEQ ID NOs: 1-8, and polypeptide sequences, SEQ

5 ID NOs: 9-10. Each sequence is identified by a sequence identification number (SEQ ID NO) and by the Incyte Clone number from which the sequence was first identified.

DESCRIPTION OF THE INVENTION

It must be noted that as used herein and in the appended claims, the singular forms

10 "a," "an," and "the" include the plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

15 **DEFINITIONS**

"NSEQ" refers generally to a polynucleotide sequence of the present invention, including SEQ ID NOs: 1-8. "PSEQ" refers generally to a polypeptide sequence of the present invention, including SEQ ID NOs: 9-10.

A " variant" refers to either a polynucleotide or a polypeptide whose sequence

20 diverges from SEQ ID NOs: 1-10 or SEQ ID NOs:9-10, respectively. Polynucleotide sequence divergence may result from mutational changes such as deletions, additions, and substitutions of one or more nucleotides; it may also occur because of differences in codon usage. Each of these types of changes may occur alone, or in combination, one or more times in a given sequence. Polypeptide variants include sequences that possess at least

25 one structural or functional characteristic of SEQ ID NOs: 9-10.

"Gene" or "gene sequence" refers to the partial or complete coding sequence of a gene. The term also refers to 5' or 3' untranslated regions. The gene may be in a sense or antisense (complementary) orientation.

"Prostate cancer-specific gene" refers to a gene sequence which has been

30 previously identified as useful in the diagnosis, treatment, prognosis, or prevention of prostate cancer. Typically, this means that the prostate cancer-specific gene is expressed at higher levels in prostate cancer tissue when compared with healthy tissue.

"Prostate cancer-associated gene" refers to a gene sequence whose expression pattern is similar to that of the prostate cancer-specific genes and which are useful in the diagnosis, treatment, prognosis, or prevention of cancer. The gene sequences can also be used in the evaluation of therapies for cancer.

5   "Substantially purified" refers to a nucleic acid or an amino acid sequence that is removed from its natural environment and is isolated or separated, and is at least about 60% free, preferably about 75% free, and most preferably about 90% free from other components with which it is naturally present.

10 **THE INVENTION**

The present invention encompasses a method for identifying biomolecules that are associated with a specific disease, regulatory pathway, subcellular compartment, cell type, tissue type, or species. In particular, the method identifies gene sequences useful in diagnosis, prognosis, treatment, prevention, and evaluation of therapies for diseases
15 associated with cell proliferation, particularly cancer, and more particularly prostate cancer.

The method entails first identifying polynucleotides that are expressed in the cDNA libraries. The polynucleotides include genes of known function, genes known to be specifically expressed in a specific disease process, subcellular compartment, cell type,
20 tissue type, or species. Additionally, the polynucleotides include genes of unknown function. The expression patterns of the known genes are then compared with those of the genes of unknown function to determine whether a specified coexpression probability threshold is met. Through this comparison, a subset of the polynucleotides having a high coexpression probability with the known genes can be identified. The high coexpression
25 probability correlates with a particular coexpression probability threshold whihc is less than 0.001, and more preferably less than 0.00001.

The polynucleotides originate from cDNA libraries derived from a variety of sources including, but not limited to, eukaryotes such as human, mouse, rat, dog, monkey, plant, and yeast and prokaryotes such as bacteria and viruses. These polynucleotides can
30 also be selected from a variety of sequence types including, but not limited to, expressed sequence tags (ESTs), assembled polynucleotide sequences, full length gene coding regions, introns, regulatory sequences, 5' untranslated regions, and 3' untranslated regions.

To have statistically significant analytical results, the polynucleotides need to be expressed in at least three cDNA libraries.

The cDNA libraries used in the coexpression analysis of the present invention can be obtained from blood vessels, heart, blood cells, cultured cells, connective tissue,

5   epithelium, islets of Langerhans, neurons, phagocytes, biliary tract, esophagus, gastrointestinal system, liver, pancreas, fetus, placenta, chromaffin system, endocrine glands, ovary, uterus, penis, prostate, seminal vesicles, testis, bone marrow, immune system, cartilage, muscles, skeleton, central nervous system, ganglia, neuroglia, neurosecretory system, peripheral nervous system, bronchus, larynx, lung, nose, pleurus,

10   ear, eye, mouth, pharynx, exocrine glands, bladder, kidney, ureter, and the like. The number of cDNA libraries selected can range from as few as 20 to greater than 10,000. Preferably, the number of the cDNA libraries is greater than 500.

In a preferred embodiment, gene sequences are assembled to reflect related sequences, such as assembled sequence fragments derived from a single transcript.

15   Assembly of the polynucleotide sequences can be performed using sequences of various types including, but not limited to, ESTs, extensions, or shotgun sequences. In a most preferred embodiment, the polynucleotide sequences are derived from human sequences that have been assembled using the algorithm disclosed in "Database and System for Storing, Comparing and Displaying Related Biomolecular Sequence Information",

20   Lincoln et al., Serial No:60/079,469, filed March 26, 1998, herein incorporated by reference.

Experimentally, differential expression of the polynucleotides can be evaluated by methods including, but not limited to, differential display by spatial immobilization or by gel electrophoresis, genome mismatch scanning, representational difference analysis, and

25   transcript imaging. Additionally, differential expression can be assessed by microarray technology. These methods may be used alone or in combination.

Genes known to be prostate cancer-specific can be selected based on the use of the genes as diagnostic or prognostic markers or as therapeutic targets for prostate cancer. Preferably, the prostate cancer-specific genes include prostate-specific antigen (PSA),

30   prostatic acid phosphatase (PAP), kallikrein, seminal plasma protein, prostate-specific tranglutaminase, and the like.

The procedure for identifying novel genes that exhibit a statistically significant

coexpression pattern with prostate cancer-specific genes is as follows. First, the presence or absence of a gene sequence in a cDNA library is defined: a gene is present in a cDNA library when at least one cDNA fragment corresponding to that gene is detected in a cDNA sample taken from the library, and a gene is absent from a library when no

5 corresponding cDNA fragment is detected in the sample.

Second, the significance of gene coexpression is evaluated using a probability method to measure a due-to-chance probability of the coexpression. The probability method can be the Fisher exact test, the chi-squared test, or the kappa test. These tests and examples of their applications are well known in the art and can be found in standard

10 statistics texts (Agresti, A. (1990) Categorical Data Analysis. New York, NY, Wiley; Rice, J. A. (1988) Mathematical Statistics and Data Analysis. Pacific Grove, CA, Wadsworth & Brooks/Cole). A Bonferroni correction (Rice, supra, page 384) can also be applied in combination with one of the probability methods for correcting statistical results of one gene versus multiple other genes. In a preferred embodiment, the due-to-chance

15 probability is measured by a Fisher exact test, and the threshold of the due-to-chance probability is set to less than 0.001, more preferably less than 0.00001.

To determine whether two genes, A and B, have similar coexpression patterns, occurrence data vectors can be generated as illustrated in Table 1, wherein a gene's presence is indicated by a one and its absence by a zero. A zero indicates that the gene

20 did not occur in the library, and a one indicates that it occurred at least once.

Table 1. Occurrence data for genes A and B

	Library 1	Library 2	Library 3	...	Library N
gene A	1	1	0	...	0
gene B	1	0	1	...	0

For a given pair of genes, the occurrence data in Table 1 can be summarized in a 2x2

30 contingency table.

Table 2. Contingency table for co-occurrences of genes A and B

	Gene A present	Gene A absent	Total
Gene B present	8	2	10
Gene B absent	2	18	20
Total	10	20	30

Table 2 presents co-occurrence data for gene A and gene B in a total of 30 libraries. Both gene A and gene B occur 10 times in the libraries. Table 2 summarizes and presents 1) the number of times gene A and B are both present in a library, 2) the number of times gene A and B are both absent in a library, 3) the number of times gene A is present while gene B is absent, and 4) the number of times gene B is present while gene A is absent. The upper left entry is the number of times the two genes co-occur in a library, and the middle right entry is the number of times neither gene occurs in a library. The off diagonal entries are the number of times one gene occurs while the other does not. Both A and B are present eight times and absent 18 times, gene A is present while gene B is absent two times, and gene B is present while gene A is absent two times. The probability ("p-value") that the above association occurs due to chance as calculated using a Fisher exact test is 0.0003. Associations are generally considered significant if a p-value is less than 0.01 (Agresti, supra; Rice, supra).

This method of estimating the probability for coexpression of two genes makes several assumptions. The method assumes that the libraries are independent and are identically sampled. However, in practical situations, the selected cDNA libraries are not entirely independent because more than one library may be obtained from a single patient or tissue, and they are not entirely identically sampled because different numbers of cDNA's may be sequenced from each library (typically ranging from 5,000 to 10,000 cDNA's per library). In addition, because a Fisher exact coexpression probability is calculated for each gene versus 41,419 other genes, a Bonferroni correction for multiple statistical tests is necessary.

Using the method of the present invention, we have identified eight novel genes that exhibit strong association, or coexpression, with known genes that are prostate cancer-

specific. These prostate cancer-specific genes include glandular kallikrein, prostate seminal protein, protate-specific antigen, and prostatic acid phosphatase. The results presented in Tables 5 to 12 show that the expression of eight novel genes have direct or indirect association with the expression of cancer-specific genes, in particular prostate

5 cancer-specific genes. Therefore, the novel genes can potentially be used in diagnosis, treatment, prognosis, or prevention of cancer, or in the evaluation of therapies for cancer. Further, the gene products of the eight novel genes are potential therapeutic proteins and targets of anti-cancer therapeutics.

Therefore, in one embodiment, the present invention encompasses a polynucleotide

10 sequence comprising the sequence of SEQ ID NOs:1-8. These eight polynucleotides are shown by the method of the present invention to have strong coexpression association with prostate cancer-specific genes and with each other. The invention also encompasses a variant of the polynucleotide sequence, its complement, or 18 consecutive nucleotides of the sequences provided in the above described sequences. Variant polynucleotide

15 sequences typically have at least about 70%, more preferably at least about 85%, and most preferably at least about 95% polynucleotide sequence identity to NSEQ.

One preferred method for identifying variants entails using NSEQ and/or PSEQ sequences to search against the GenBank primate (pri), rodent (rod), and mammalian (mam), vertebrate (vrtp), and eukaryote (eukp) databases, SwissProt, BLOCKS (Bairoch,

20 A. et al. (1997) Nucleic Acids Res. 25:217-221), PFAM, and other databases that contain previously identified and annotated motifs, sequences, and gene functions. Methods that search for primary sequence patterns with secondary structure gap penalties (Smith, T. et al. (1992) Protein Engineering 5:35-51) as well as algorithms such as BLAST (Basic Local Alignment Search Tool; Altschul, S.F. (1993) J. Mol. Evol 36:290-300; and Altschul et al.

25 (1990) J. Mol. Biol. 215:403-410), BLOCKS (Henikoff S. and Henikoff G.J. (1991) Nucleic Acids Research 19:6565-6572), Hidden Markov Models (HMM; Eddy, S.R. (1996) Cur. Opin. Str. Biol. 6:361-365; and Sonnhammer, E.L.L. et al. (1997) Proteins 28:405-420), and the like, can be used to manipulate and analyze nucleotide and amino acid sequences. These databases, algorithms and other methods are well known in the art

30 and are described in Ausubel, F.M. et al. (1997; Short Protocols in Molecular Biology, John Wiley & Sons, New York , NY)and in Meyers, R.A. (1995; Molecular Biology and Biotechnology, Wiley VCH, Inc, New York, NY, p 856-853).

Also encompassed by the invention are polynucleotide sequences that are capable
of hybridizing to SEQ ID NOs:1-8, and fragments thereof under stringent conditions.
Stringent conditions can be defined by salt concentration, temperature, and other
chemicals and conditions well known in the art. In particular, stringency can be increased
5   by reducing the concentration of salt, or raising the hybridization temperature.

For example, stringent salt concentration will ordinarily be less than about 750 mM
NaCl and 75 mM trisodium citrate, preferably less than about 500 mM NaCl and 50 mM
trisodium citrate, and most preferably less than about 250 mM NaCl and 25 mM trisodium
citrate. Stringent temperature conditions will ordinarily include temperatures of at least
10   about 30°C, more preferably of at least about 37°C, and most preferably of at least about
42°C. Varying additional parameters, such as hybridization time, the concentration of
detergent (sodium dodecyl sulfate, SDS) or solvent (formamide), and the inclusion or
exclusion of carrier DNA, are well known to those skilled in the art. Additional variations
on these conditions will be readily apparent to those skilled in the art (Wahl, G.M. and
15   S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods
Enzymol. 152:507-511; Ausubel, F.M. et al. (1997) Short Protocols in Molecular Biology,
John Wiley & Sons, New York, NY; and Sambrook, J. et al. (1989) Molecular Cloning, A
Laboratory Manual, Cold Spring Harbor Press, Plainview, NY).

NSEQ or the polynucleotide sequences encoding PSEQ can be extended utilizing a
20   partial nucleotide sequence and employing various PCR-based methods known in the art
to detect upstream sequences, such as promoters and regulatory elements. (See, e.g.,
Dieffenbach, C.W. and G.S. Dveksler (1995; PCR Primer, a Laboratory Manual, Cold
Spring Harbor Press, Plainview, NY, pp.1-5; Sarkar, G. (1993; PCR Methods Applic.
2:318-322); Triglia, T. et al. (1988; Nucleic Acids Res. 16:8186); Lagerstrom, M. et al.
25   (1991; PCR Methods Applic. 1:111-119); and Parker, J.D. et al. (1991; Nucleic Acids Res.
19:3055-306). Additionally, one may use PCR, nested primers, and
PROMOTERFINDER libraries to walk genomic DNA (Clontech, Palo Alto, CA). This
procedure avoids the need to screen libraries and is useful in finding intron/exon junctions.
For all PCR-based methods, primers may be designed using commercially available
30   software, such as OLIGO 4.06 Primer Analysis software (National Biosciences Inc.,
Plymouth MN) or another appropriate program, to be about 18 to 30 nucleotides in length,
to have a GC content of about 50% or more, and to anneal to the template at temperatures

of about 68°C to 72°C.

In another aspect of the invention, NSEQ or the polynucleotide sequences encoding PSEQ can be cloned in recombinant DNA molecules that direct expression of PSEQ or the polypeptides encoded by NSEQ, or structural or functional fragments thereof,
5 in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express the polypeptides of PSEQ or the polypeptides encoded by NSEQ. The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter the nucletide
10 sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction
15 sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

In order to express a biologically active polypeptide encoded by NSEQ, NSEQ or the polynucleotide sequences encoding PSEQ, or derivatives thereof, may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for
20 transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in NSEQ or polynucleotide sequences encoding PSEQ. Methods which are well known to those skilled in the art may be used to construct expression vectors containing NSEQ or
25 polynucleotide sequences encoding PSEQ and appropriate transcriptional and translational control elements. These methods include _in vitro_ recombinant DNA techniques, synthetic techniques, and _in vivo_ genetic recombination. (See, e.g., Sambrook (_supra_) and Ausubel, (_supra_).

A variety of expression vector/host cell systems may be utilized to contain and
30 express NSEQ or polynucleotide sequences encoding PSEQ. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression

vectors; insect cell systems infected with viral expression vectors (baculovirus); plant cell systems transformed with viral expression vectors, cauliflower mosaic virus (CaMV) or tobacco mosaic virus (TMV), or with bacterial expression vectors (Ti or pBR322 plasmids); or animal cell systems. The invention is not limited by the host cell employed.

5   For long term production of recombinant proteins in mammalian systems, stable expression of a polypeptide encoded by NSEQ in cell lines is preferred. For example, NSEQ or sequences encoding PSEQ can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector.

10   In general, host cells that contain NSEQ and that express PSEQ may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein

15   sequences. Immunological methods for detecting and measuring the expression of PSEQ using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS).

Host cells transformed with NSEQ or polynucleotide sequences encoding PSEQ
20   may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides of NSEQ or polynucleotides encoding PSEQ may be designed to contain signal sequences which direct

25   secretion of PSEQ or polypeptides encoded by NSEQ through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation,

30   glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and

characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38), are available from the American Type Culture Collection (ATCC, Bethesda, MD) and may be chosen to ensure the correct modification and processing of the foreign protein.

5   In another embodiment of the invention, natural, modified, or recombinant NSEQ or nucleic acid sequences encoding PSEQ are ligated to a heterologous sequence resulting in translation of a fusion protein containing heterologous protein moieties in any of the aforementioned host systems. Such heterologous protein moieties facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, 10   but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, *c-myc*, hemagglutinin (HA) and monoclonal antibody epitopes..

In another embodiment, NSEQ or sequences encoding PSEQ are synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, 15   M.H. et al. (1980) Nucl. Acids Res. Symp. Ser. 215-223; Horn, T. et al. (1980) Nucl. Acids Res. Symp. Ser. 225-232; and Ausubel, supra). Alternatively, PSEQ or a polypeptide sequence encoded by NSEQ itself, or a fragment thereof, may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solid-phase techniques (Roberge, J.Y. et al. (1995) Science 269:202-204). Automated 20   synthesis may be achieved using the ABI 431A Peptide Synthesizer (Perkin Elmer). Additionally, PSEQ or the amino acid sequence encoded by NSEQ, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a polypeptide variant.

In another embodiment, the invention entails a substantially purified polypeptide 25   comprising the amino acid sequence selected from the group consisting of SEQ ID NO:9, SEQ ID NO:10, or fragments thereof. SEQ ID NO:9 is encoded by SEQ ID NO:4 and is a potential transmembrane protein which interacts with a cell surface receptor. SEQ ID NO:10 is encoded by SEQ ID NO:8 and has potential sequence homology with a family of GPI-linked cell-surface glycoproteins, Ly-6/u-PAR.

30

DIAGNOSTICS and THERAPEUTICS

The sequences of the these genes can be used in diagnosis, prognosis, treatment,

prevention, and evaluation of therapies for diseases associated with cell proliferation, particularly cancer, and more particularly prostate cancer. Further, the amino acid sequences encoded by the novel genes are potential therapeutic proteins and targets of anti-cancer therapeutics.

5   In one preferred embodiment, the polynucleotide sequences of NSEQ or the polynucleotides encoding PSEQ are used for diagnostic purposes to determine the absence, presence, and excess expression of PSEQ, and to monitor regulation of the levels of mRNA or the polypeptides encoded by NSEQ during therapeutic intervention. The polynucleotides may be at least 18 nucleotides long, complementary RNA and DNA

10   molecules, branched nucleic acids, and peptide nucleic acids (PNAs). Alternatively, the polynucleotides are used to detect and quantitate gene expression in samples in which expression of PSEQ or the polypeptides encoded by NSEQ are correlated with disease. Additionally, NSEQ or the polynucleotides encoding PSEQ can be used to detect genetic polymorphisms associated with a disease. These polymorphisms may be detected at the

15   transcript cDNA or genomic level.

        The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification (maximal, high, intermediate, or low), will determine whether the probe identifies only naturally occurring sequences encoding

20   PSEQ, allelic variants, or related sequences.

        Probes may also be used for the detection of related sequences, and should preferably have at least 50% sequence identity to any of the NSEQ or PSEQ-encoding sequences.

        Means for producing specific hybridization probes for DNAs encoding PSEQ

25   include the cloning of NSEQ or polynucleotide sequences encoding PSEQ into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such

30   as ^{32}P or ^{35}S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, by fluorescent labels and the like. The polynucleotide sequences encoding PSEQ may be used in Southern or northern analysis, dot blot, or other

membrane-based technologies; in PCR technologies; and in microarrays utilizing fluids or tissues from patients to detect altered PSEQ expression. Such qualitative or quantitative methods are well known in the art.

NSEQ or the nucleotide sequences encoding PSEQ can be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantitated and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to the standard value then the presence of altered levels of nucleotide sequences of NSEQ and those encoding PSEQ in the sample indicates the presence of the associated disease. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

Once the presence of a disease is established and a treatment protocol is initiated, hybridization or amplification assays can be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

The polynucleotides may be used for the diagnosis of a variety of diseases associated with cell proliferation including cancer such as adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus.

Alternatively, the polynucleotides may be used as targets in a microarray. The microarray can be used to monitor the expression level of large numbers of genes simultaneously and to identify splice variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disease, to diagnose a disease, and to develop and monitor the activities of therapeutic agents.

In yet another alternative, polynucleotides may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Fluorescent in situ hybridization (FISH) may be correlated with other physical chromosome mapping

techniques and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, R.A. (ed.) Molecular Biology and Biotechnology, VCH Publishers New York, NY, pp. 965-968).

In another embodiment, antibodies which specifically bind PSEQ may be used for
5   the diagnosis of diseases characterized by the over-or-underexpression of PSEQ or polypeptides encoded by NSEQ. Alternatively, one may use competitive drug screening assays in which neutralizing antibodies capable of binding PSEQ or the polypeptides encoded by NSEQ specifically compete with a test compound for binding the polypeptides. In this manner, antibodies can be used to detect the presence of any peptide
10 which shares one or more antigenic determinants with PSEQ or the polypeptides encoded by NSEQ. Diagnostic assays for PSEQ or the polypeptides encoded by NSEQ include methods which utilize the antibody and a label to detect PSEQ or the polypeptided encoded by NSEQ in human body fluids or in extracts of cells or tissues. A variety of protocols for measuring PSEQ or the polypeptides encoded by NSEQ, including ELISAs,
15 RIAs, and FACS, are well known in the art and provide a basis for diagnosing altered or abnormal levels of the expression of PSEQ or the polypeptides encoded by NSEQ. Normal or standard values for PSEQ expression are established by combining body fluids or cell extracts taken from normal subjects, preferably human, with antibody to PSEQ or a polypeptide encoded by NSEQ under conditions suitable for complex formation The
20 amount of standard complex formation may be quantitated by various methods, preferably by photometric means. Quantities of PSEQ or the polypeptides encoded by NSEQ expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing or monitoring disease.

25   In another aspect, the polynucleotides and polypeptides of the present invention can be employed for treatment or the monitoring of therapeutic treatments for cancers. The polynucleotides of NSEQ or those encoding PSEQ, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, the complement of the polynucleotides of NSEQ or those encoding PSEQ may be used in situations in which it
30 would be desirable to block the transcription or translation of the mRNA.

Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide

sequences to the targeted organ, tissue, or cell population. Methods which are well known to those skilled in the art can be used to construct vectors to express nucleic acid sequences complementary to the polynucleotides encoding PSEQ. (See, e.g., Sambrook, supra; and Ausubel, supra.)

5   Genes having polynucleotide sequences of NSEQ or those encoding PSEQ can be turned off by transforming a cell or tissue with expression vectors which express high levels of a polynucleotide, or fragment thereof, encoding PSEQ. Such constructs may be used to introduce untranslatable sense or antisense sequences into a cell. Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10

10   from the start site, are preferred. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber,

15   B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing Co., Mt. Kisco, NY, pp. 163-177.) Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at

20   the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine,

25   thymine, and uridine which are not as easily recognized by endogenous endonucleases.

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or

30   by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nature Biotechnology 15:462-466.)

Further, an antagonist or antibody of a polypeptide of PSEQ or encoded by NSEQ

may be administered to a subject to treat or prevent a cancer associated with increased expression or activity of PSEQ. An antibody which specifically binds the polypeptide may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissue which express the the polypeptide.

5   Antibodies to PSEQ or polypeptides encoded by NSEQ may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are especially preferred for therapeutic use. Monoclonal

10   antibodies to PSEQ may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. In addition, techniques developed for the production of chimeric antibodies can be used. (See, for example, Molecular Biology and

15   Biotechnology, R.A. Myers, ed.,(1995)John Wiley & Sons, Inc., New York, NY). Alternatively, techniques described for the production of single chain antibodies may be employed. Antibody fragments which contain specific binding sites for PSEQ or the polypeptide sequences encoded by NSEQ may also be generated.

          Various immunoassays may be used for screening to identify antibodies having the

20   desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art.

          Yet further, an agonist of a polypeptide of PSEQ or that encoded by NSEQ may be administered to a subject to treat or prevent a cancer associated with decreased expression

25   or activity of the polypeptide.

          An additional aspect of the invention relates to the administration of a pharmaceutical or sterile composition, in conjunction with a pharmaceutically acceptable carrier, for any of the therapeutic effects discussed above. Such pharmaceutical compositions may consist of polypeptides of PSEQ or those encoded by NSEQ, antibodies

30   to the polypeptides, and mimetics, agonists, antagonists, or inhibitors of the polypeptides. The compositions may be administered alone or in combination with at least one other agent, such as a stabilizing compound, which may be administered in any sterile,

biocompatible pharmaceutical carrier including, but not limited to, saline, buffered saline, dextrose, and water. The compositions may be administered to a patient alone, or in combination with other agents, drugs, or hormones.

The pharmaceutical compositions utilized in this invention may be administered by
5 any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically-acceptable carriers comprising excipients and auxiliaries
10 which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing Co., Easton, PA).

For any compound, the therapeutically effective dose can be estimated initially
15 either in cell culture assays, e.g., of neoplastic cells or in animal models such as mice, rats, rabbits, dogs, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for
20 example, polypeptides of PSEQ or those encoded by NSEQ, or fragments thereof, antibodies of the polypeptides, and agonists, antagonists or inhibitors of the polypeptides, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED_{50} (the dose therapeutically effective in 50% of the
25 population) or LD_{50} (the dose lethal to 50% of the population) statistics.

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as dogs, cats, cows, horses, rabbits, monkeys, and most preferably, humans.

30

EXAMPLES

It is understood that this invention is not limited to the particular methodology, protocols, and reagents described, as these may vary. It is also understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims. The examples below are provide to illustrate the subject invention and are not included for the purpose of limiting the invention.

I. PROSTUT05 cDNA Library Construction

For purposes of example, the preparation of the PROSTUT05 library is described. The PROSTUT05 cDNA library was constructed using polyA RNA isolated from prostate tumor tissue removed from a 69-year-old Caucasian male during a radical prostatectomy. Pathology indicated adenocarcinoma (Gleason grade 3+4) involving the right side peripherally. The tumor invaded the capsule but did not extend beyond it; perineural invasion was present. Adenofibromatous hyperplasia was also present. The right seminal vesicle was involved with tumor. The patient presented with elevated prostate specific antigen (PSA). Patient history included partial colectomy, and tobacco use. Family history included congestive heart failure, multiple myeloma, hyperlipidemia, and rheumatoid arthritis.

The frozen tissue was homogenized and lysed using a Brinkmann Homogenizer Polytron PT-3000 (Brinkmann Instruments, Westbury, NJ) in guanidinium isothiocyanate solution. The lysate was centrifuged over a 5.7 M CsCl cushion using an Beckman SW28 rotor in a Beckman L8-70M Ultracentrifuge (Beckman Instruments) for 18 hours at 25,000 rpm at ambient temperature. The RNA was extracted with acid phenol pH 4.7, precipitated using 0.3 M sodium acetate and 2.5 volumes of ethanol, resuspended in RNAse-free water, and treated with DNase at 37°C. mRNA extraction and precipitation were repeated as before. The mRNA was isolated with the Qiagen Oligotex kit (QIAGEN, Chatsworth, CA) and used to construct the cDNA library.

The mRNA was handled according to the recommended protocols in the SuperScript Plasmid System for cDNA synthesis and plasmid cloning (GIBCO/BRL). The cDNAs were fractionated on a Sepharose CL4B column (Pharmacia), and those cDNAs exceeding 400 bp were ligated into pSport I. The plasmid pSport I was subsequently transformed into DH5α™ competent cells (LifeTechnologies, Gaithersburg,

MD).

II. Isolation and Sequencing of cDNA Clones

Plasmid DNA was released from the cells and purified using the REAL Prep 96
5 plasmid kit (QIAGEN). This kit enabled the simultaneous purification of 96 samples in a
96-well block using multi-channel reagent dispensers. The recommended protocol was
employed except for the following changes: 1) the bacteria were cultured in 1 ml of sterile
Terrific Broth (LifeTechnologies) with carbenicillin at 25 mg/L and glycerol at 0.4%; 2)
after inoculation, the cultures were incubated for 19 hours and at the end of incubation, the
10 cells were lysed with 0.3 ml of lysis buffer; and 3) following isopropanol precipitation, the
plasmid DNA pellet was resuspended in 0.1 ml of distilled water. After the last step in the
protocol, samples were transferred to a 96-well block for storage at 4° C.

The cDNAs were prepared and sequenced by the method of Sanger et al. (1975, J.
Mol. Biol. 94:441f), using a Hamilton Micro Lab 2200 (Hamilton, Reno, NV) in
15 combination with Peltier Thermal Cyclers (PTC200 from MJ Research, Watertown, MA)
and Applied Biosystems 373 and 377 DNA Sequencing Systems.

III. Selection, Assembly, and Characterization of Sequences

The sequences used for coexpression analysis were assembled from EST
20 sequences, 5' and 3' longread sequences, and full length coding sequences. Selected
assembled sequences were expressed in at least three cDNA libraries.

The assembly process is described as follows. EST sequence chromatograms were
processed and verified. Quality scores were obtained using PHRED (Ewing, B. et al.
(1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194).
25 Then the edited sequences were loaded into a relational database management system
(RDBMS). The EST sequences were clustered into an initial set of bins using BLAST
with a product score of 50. All clusters of two or more sequences were created as bins.
The overlapping sequences represented in a bin correspond to the sequence of a
transcribed gene.

30   Assembly of the component sequences within each bin was performed using a
modification of Phrap, a publicly available program for assembling DNA fragments
(Green, P., University of Washington, Seattle, WA). Bins that showed 82% identity from a

-22-

local pair-wise alignment between any of the consensus sequences were merged.

Bins were annotated by screening the consensus sequence in each bin against public databases, such as gbpri and genpept from NCBI. The annotation process involved a FASTn screen against the gbpri database in GenBank. Those hits with a percent identity

5   of greater than or equal to 70% and an alignment length of greater than or equal to 100 base pairs were recorded as homolog hits. The residual unannotated sequences were screened by FASTx against genpept. Those hits with an E value of less than or equal to 10^{-8} are recorded as homolog hits.

Sequences were then reclustered using BLASTn and Cross-Match, a program for

10 rapid protein and nucleic acid sequence comparison and database search (Green, P., University of Washington, Seattle, WA), sequentially. Any BLAST alignment between a sequence and a consensus sequence with a score greater than 150 was realigned using cross-match. The sequence was added to the bin whose consensus sequence gave the highest Smith-Waterman score amongst local alignments with at least 82% identity. Non-

15 matching sequences created new bins. The assembly and consensus generation processes were performed for the new bins.

IV.   Co-expression Analyses of Known Prostate Cancer-Specific Genes

Five known prostate cancer-specific genes were selected to test the validity of the

20 coexpression analysis method of the present invention in identifying genes that are closely associated with prostate cancer. The five known genes were prostate-specific antigen, glandular kallikrein, prostate seminal protein, prostatic acid phosphatase, and prostate transglutaminase. As shown, the method successfully identified the strong association of the known genes among themselves, indicating that the coexpression analysis method of

25 the present invention was effective in identifying genes that are closely associated with prostate cancer.

Table 4 shows the top ten genes that were most closely associated with a known prostate cancer-specific gene. These genes are presented along with their p-values. The column headings have the following meanings:

30

P-value   The probability that the observed number of
co-occurrences is due to chance using the Fisher

exact method.
Co-expressed Gene A gene that shows significant co-expression with   the
target.

No. Occur   The number of libraries in which the associated   gene
5 occurs.

No. Co-occur   The number of libraries in which both the target   gene
and the co-expressed gene occur.

No. Target Only   The number of libraries in which **only** the target   gene
occurs.

10 No. Gene Only   The number of libraries in which **only** the
associated gene occurs.

No. Neither Occur The number of libraries in which neither the   target
gene nor the associated gene occur.

15

Table 4. Co-expression results for prostate-specific antigen

P-value	Co-expressed Gene	No. Occur	No. Co-occur	No. Target Only	No. Gene Only	No. Neither Occur
1.53E-31	Glandular kallikrein	29	26	12	3	481
1.65E-26	1816556	24	22	16	2	482
7.48E-25	1864683	40	26	12	14	470
8.12E-25	1344875	29	23	15	6	478
3.38E-24	Prostate seminal protein	30	23	15	7	477
1.89E-23	Prostatic acid phosphatase	35	24	14	11	473
6.87E-18	1651189	28	19	19	9	475
9.01E-18	Prostate trans-glutaminase	14	14	24	0	484
4.61E-14	1646118	93	27	11	66	418
1.58E-13	Human neuropeptide Y (NPY) mRNA	27	16	22	11	473

As a target, prostate-specific antigen occurred in 38 of 522 cDNA libraries studied, and showed strong coexpression with glandular kallikrein, prostate seminal protein, prostatic acid phosphatase, and prostate transglutaminase. The target also showed strong association with the human neuropeptide tyrosine (NPY) mRNA. In addition, four of the

5   top ten genes that showed strong association with the target were novel Incyte assembled genes: 1816556, 1864683, 1344875, 1651189, and 1646118. These results are shown in Table 4 with association probability in the range of 1.58E-13 to 1.53E-31.

Similar results were observed when the other four known prostate cancer-specific genes, glandular kallikrein, prostate seminal protein, prostatic acid phosphatase, and

10   prostate transglutaminase, were taken as target genes. These target genes were also found to be strongly associated with several known genes, many of which are cancer-related. The cancer-related genes are human neuropeptide tyrosine (NPY), human serine protease encoded by TMPRSS2, sorbitol dehydrogenase isozyme, human Zn-alpha-2-glycoprotein, and MAT-8.

15   NPY was first isolated from a human pheochromocytoma tumor (Minth, C. D. et al. (1984) Proc Natl Acad Sci 81(14): 4577-4581) and was reported to be involved in prostate cancer (Rogatnick, L. A. et al. (1990) Proc West Pharmacol Soc 33: 47-53; Mack, D., G. et al. (1997) Eur J Cancer 33:317-318). The TMPRSS2 gene was identified as a gene that encodes a serine protease domain specific for cleavage at Arg or Lys residues

20   (Paoloni-Giacobino, A. et al. (1997) Genomics 44:309-320). The protease activity of TMPRSS2 is similar to that of PSA and kallikrein, both human prostate cancer-specific genes. Sorbitol dehydrogenase isozyme has been used as a marker for male reproductive tissue, including the prostate (Holmes, R. S. et al. (1978) J Exp Zool 206: 279-88). Significant activity of the enzyme accompanies damage to reproductive tissue.

25   Zn-alpha-2-glycoprotein is a secreted protein identified in hormone-responsive breast carcinomas (Freije, J. P. et al. (1993)Genomics 18:575-87) and was proposed as a marker for breast carcinomas (Lopez-Boado, Y. S. et al. (1994) Breast Cancer Res Treat 29: 247-58). It was shown to be prognostic for a 5-year breast cancer survival (Hurlimann, J. and G. van Melle (1991) Am J Clin Pathol 95:835-43) and was reported to show

30   differential expression in prostate carcinoma (Gagnon, S. et al. (1990) Am J Pathol 136: 1147-52). MAT-8 was first identified in murine breast tumors and subsequently in primary human breast tumors and cell lines (Morrison, B. W. et al. (1995) J Biol Chem

270: 2176-82). It was shown to be a marker for progression of breast cancer (Schiemann, S. et al. (1998) Clin Exp Metastasis 16:129-39). A relation to prostate cancer has not been previously reported.

5 **V.   Identification of Novel Prostate Cancer-Associated Genes**

Using the coexpression analysis, we have identified eight novel genes that show strong association with prostate cancer from a total of 41,419 assembled gene sequences. The degree of association was measured by probability values and has a cutoff of p value less than 0.00001. This was followed by annotation and literature searches to insure that the genes that passed the probability test have strong association with known prostate cancer-specific genes. This process was reiterated so that the initial 41419 genes were reduced to the final eight prostate cancer-associated genes. Details of identification for the eight novel prostate cancer-associated genes are presented in Tables 5 to 12. These tables show the ten genes that were most closely associated for each target novel gene as measured by coexpression using the Fisher exact test. The column headings have the same meanings as in Example IV.

Table 5. Co-expression results for Incyte gene 842349

P-value	Co-expressed Gene	No. Occur	No. Co-occur	No. Target Only	No. Gene Only	No. Neither Occur
4.23E-11	Glandular kallikrein	29	17	38	12	455
1.29E-09	Prostate seminal protein	30	16	39	14	453
3.62E-09	1816556	24	14	41	10	457
8.56E-09	1344875	29	15	40	14	453
1.09E-08	TMPRSS2-encoded serine protease	74	24	31	50	417
1.19E-08	Prostate-specific antigen	38	17	38	21	446

2.45E-08	Prostatic acid phosphatase	35	16	39	19	448
2.91E-08	Human neuropeptide Y (NPY) mRNA	27	14	41	13	454
2.92E-08	1697453	96	27	28	69	398
3.14E-08	1864683	40	17	38	23	444

Incyte gene 842349 occurred in 55 of 522 cDNA libraries studied and showed strong co-expression with several of the known prostate cancer-specific genes, including glandular kallikrein, prostate seminal protein, prostate-specific antigen, and prostatic acid phosphatase, as shown in Table 9. 842349 also showed strong association with a human TMPRSS2-encoded serine protease. The serine protease was shown to be strongly associated with prostate cancer-specific prostatic acid phosphatase in Example IV. Further, 842349 showed strong association with four novel Incyte genes, 1816556, 1344875, 1697453, and 1864683. These results are consistent with the notion that 842349 is associated with prostate cancer; and 842349 may be functionally or regulatorily associated with at least four novel Incyte genes.

Table 6. Co-expression results for 1682557

P-value	Co-expressed Gene	No. Occur	No. Co-occur	No. Target Only	No. Gene Only	No. Neither Occur
1.34E-07	1816556	24	5	0	19	498
3.40E-07	Human nicotinic acetylcholine receptor A	10	4	1	6	511
3.75E-07	Glandular kallikrein	29	5	0	24	493
3.75E-07	1344875	29	5	0	24	493
1.02E-06	Prostatic acid phosphatase	35	5	0	30	487

1.58E-06	Prostate-specific antigen	38	5	0	33	484
2.08E-06	1864683	40	5	0	35	482
4.22E-06	1685804	5	3	2	2	515
9.53E-06	3096181	21	4	1	17	500
2.34E-05	1794279	8	3	2	5	512

Incyte gene 1682557 occurred in 5 of 522 cDNA libraries studied and showed strong coexpression with several of the known prostate cancer-specific genes, such as glandular kallikrein, prostatic acid phosphatase, and prostate-specific antigen, as shown in Table 10. 1682557 also exhibited strong coexpression with the human nicotinic acetylcholine receptor A, a neurotransmitter which is a ligand-gated cation channel and causes rapid depolarization in postsynaptic cells. Further, Table 10 shows that 842349 has strong association with five novel Incyte genes, 1816556, 1344875, 1697453, 1864683, and 1794279. These results are consistent with the notion that 1682557 is associated with prostate cancer; and 1682557 may be functionally or regulatorily associated with at least five novel Incyte genes.

Table 7. Co-expression results for 1816556

P-value	Co-expressed Gene	No. Occur	No. Co-Occur	No. Target Only	No. Gene Only	No. Neither Occur
1.20E-30	Glandular kallikrein	29	22	2	7	491
1.11E-27	Prostatic acid phosphatase	35	22	2	13	485
1.65E-26	Prostate-specific antigen	38	22	2	16	482
1.55E-25	1344875	29	20	4	9	489
1.55E-23	1864683	40	21	3	19	479
8.19E-23	Prostate seminal protein	30	19	5	11	487

1.03E-16	Human gene for ZN-alpha-2-glycopr otein	86	22	2	64	434
3.13E-16	Prostate trans-glutaminase	14	12	12	2	496
1.08E-15	1651189	28	15	9	13	485
4.81E-15	2819055	44	17	7	27	471

Incyte gene 1816556 occurred in 24 of 522 cDNA libraries studied and showed

strong co-expression with several of the known prostate cancer-specific genes, such as

glandular kallikrein, prostatic acid phosphatase, and prostate-specific antigen, prostate

seminal protein, and prostate transglutaminase, as shown in Table 11. 1816556 also

exhibited strong association with a human gene for ZN-alpha-2-glycoprotein which was

shown in Example IV to be strongly associated with a prostate cancer-specific gene

encoding prostatic transglutaminase. Further, 1816556 showed strong co-expression with

four novel Incyte genes, 1344875, 1864683, 1651189, and 2819055. These results are

consistent with the notion that 1816556 is associated with prostate cancer; and 1816556

may be functionally or regulatorily associated with at least four novel Incyte genes.

Table 8. Co-expression results for 1864683

P-value	Co-expressed Gene	No. Occur	No. Co-Occur	No. Target Only	No. Gene Only	No. Neither Occur
7.48E-25	Prostate-specific antigen	38	26	14	12	470
4.55E-24	1344875	29	23	17	6	476
4.55E-24	Glandular kallikrein	29	23	17	6	476
1.55E-23	1816556	24	21	19	3	479
1.43E-21	Prostate seminal protein	30	22	18	8	474

6.52E-21	Prostatic acid phosphatase	35	23	17	12	470
5.43E-15	Prostate trans-glutaminase	14	13	27	1	481
6.38E-15	TMPRSS2-encoded serine protease	74	26	14	48	434
3.41E-14	1651189	28	17	23	11	471
1.47E-13	2819055	44	20	20	24	458

Incyte gene 1864683 occurred in 40 of 522 cDNA libraries studied and showed strong co-expression with several of the known prostate cancer-specific genes, such as prostate-specific antigen, glandular kallikrein, prostate seminal protein, prostatic acid phosphatase, and prostate transglutaminase, as shown in Table 8. 1864683 also exhibited strong association with a human TMPRSS2-encoded serine protease shown in Example IV to be strongly associated with prostate cancer-specific gene encoding prostatic acid phosphatase. Further, 1864683 showed strong coexpression with four novel Incyte genes, 1344875, 1816556, 1651189, and 2819055. These results are consistent with the notion that 1864683 is associated with prostate cancer; and 1864683 may be functionally or regulatorily associated with at least four novel Incyte genes.

Table 9. Co-expression results for 2187866

P-value	Co-expressed Gene	No. Occur	No. Co-Occur	No. Target Only	No. Gene Only	No. Neither Occur
9.11E-11	Prostatic acid phosphatase	35	9	1	26	486
2.42E-10	1816556	24	8	2	16	496
1.38E-09	Glandular kallikrein	29	8	2	21	491
1.38E-09	1344875	29	8	2	21	491

1.53E-08	Prostate-specific antigen	38	8	2	30	482
2.38E-08	1864683	40	8	2	32	480
3.69E-08	Human LPAP gene	98	10	0	88	424
5.41E-08	2819055	44	8	2	36	476
1.32E-07	TMPRSS2-encoded serine protease	74	9	1	65	447
2.34E-06	843197	7	4	6	3	509

Incyte gene 2187866 occurred in 10 of 522 cDNA libraries and showed strong
association with several of the known prostate cancer-specific genes, such as prostatic acid
phosphatase, glandular kallikrein, and prostate-specific antigen, as shown in Table 13.
2187866 also exhibited strong association with a human lymphocyte phosphatase-
associated phosphoprotein (LPAP) gene and a human TMPRSS2-encoded serine protease.
LPAP is a 32 kDa protein that non-covalently binds tyrosine phosphatase CD45 (Bruyns,
E., et al. (1998)Int Immunol 10: 185-94; Bruyns, E., A et al. (1996) Genomics 38: 79-83).
As specified in Example IV, TMPRSS2-encoded serine protease was associated with a
prostate cancer-specific gene encoding prostatic acid phosphatase. Further, 2187866
exhibited association with five novel Incyte genes, 1816556, 1344875, 1864683, 2819055,
and 843197. These results are consistent with the notion that 2187866 is associated with
prostate cancer; and 2187866 may be functionally or regulatorily associated with at least
five novel Incyte genes.

Table 10. Co-expression results for 3096181

P-value	Co-expressed Gene	No. Occur	No. Co-Occur	No. Target Only	No. Gene Only	No. Neither Occur
3.69E-13	1344875	29	13	8	16	485
4.96E-10	Glandular kallikrein	29	11	10	18	483

6.84E-10	Prostate-specific antigen	38	12	9	26	475
8.49E-10	Human gene for ZN-alpha-2-glycoprotein	86	16	5	70	431
1.38E-09	1816556	24	10	11	14	487
5.35E-09	Prostatic acid phosphatase	35	11	10	24	477
1.87E-08	Prostate seminal protein	30	10	11	20	481
2.70E-08	1864683	40	11	10	29	472
1.26E-07	Human T-cell receptor gamma chain	27	9	12	18	483
1.67E-07	Prostate trans-glutaminase	14	7	14	7	494

Incyte gene 3096181 occurred in 21 of 522 libraries studied and showed strong co-expression with several of the known prostate cancer-specific genes, such as glandular kallikrein, prostate-specific antigen, prostatic acid phosphatase, prostate seminal protein, and prostate transglutaminase, as shown in Table 14. 3096181 also exhibited strong coexpression with a human gene for ZN-alpha-2-glycoprotein and human T-cell receptor gamma chain. As specified in Example IV, the human gene for ZN-alpha-2-glycoprotein was associated with a prostate cancer-specific gene encoding prostatic transglutaminase. ZN-alpha-2-glycoprotein itself was identified in hormone-responsive breast carcinomas (Freije et al., supra). Human T-cell receptor gamma/delta is expressed in tumor-infiltrating lymphocytes in breast carcinoma patients (Alam, S.M. et al. (1992) Immunol Lett 31:279-283) and in malignant lymphoma presenting with hepatosplenic disease (Farcet, J. P. et al. (1990) Blood 75: 2213-2219). T-cell receptor gamma positive cells are over-expressed in cancer patients (Seki, S. et al. (1990) J Clin Invest 86: 409-15). Further, 3096181 exhibited coexpression with three novel Incyte genes, 1344875, 1816556, and 1864683. These results are consistent with the notion that 3096181 is

associated with prostate cancer; and 3096181 may be functionally or regulationally associated with at least three novel Incyte genes.

Table 11. Co-expression results for 3360806

P-value	Co-expressed Gene	No. Occur	No. Co-Occur	No. Target Only	No. Gene Only	No. Neither Occur
1.98E-11	Prostate-specific antigen	38	16	18	22	466
6.51E-10	1651189	28	13	21	15	473
8.24E-10	1864683	40	15	19	25	463
1.88E-08	1344875	29	12	22	17	471
1.88E-08	Glandular kallikrein	29	12	22	17	471
3.00E-08	Prostate seminal protein	30	12	22	18	470
5.10E-08	Human gene for ZN-alpha-2-glycoprotein	86	19	15	67	421
3.19E-07	Prostate trans-glutaminase	14	8	26	6	482
3.66E-07	1816556	24	10	24	14	474
6.56E-07	1685861	133	22	12	111	377

Incyte gene 3360806 occurred in 34 of 522 cDNA libraries and showed strong co-expression with several of the known prostate cancer-specific genes, such as prostate-specific antigen, glandular kallikrein, prostate seminal protein, and prostate transglutaminase, as shown in Table 15. 3360806 also exhibited strong co-expression with a human gene for ZN-alpha-2-glycoprotein shown in Example IV to be associated with a prostate cancer-specific gene encoding prostatic transglutaminase. ZN-alpha-2-glycoprotein itself was also found in hormone-responsive breast carcinomas (Freije et al., supra). Further, 3360806 showed association with five novel Incyte genes, 1651189, 1864683, 1344875, 1816556, and 1685861. These results are consistent with the

notion that 3360806 is associated with prostate cancer; and 3360806 may be functionally or regulationally associated with at least four novel Incyte genes.

Table 12. Co-expression results for 3458076

P-value	Co-expressed Gene	No. Occur	No. Co-Occur	No. Target Only	No. Gene Only	No. Neither Occur
3.35E-08	1816556	24	6	1	18	497
1.17E-07	1344875	29	6	1	23	492
1.17E-07	Glandular kallikrein	29	6	1	23	492
1.46E-07	Prostate seminal protein	30	6	1	24	491
3.96E-07	Prostatic acid phosphatase	35	6	1	29	486
6.70E-07	Prostate-specific antigen	38	6	1	32	483
9.29E-07	1864683	40	6	1	34	481
6.04E-06	1651189	28	5	2	23	492
1.09E-05	Prostate trans-glutaminase	14	4	3	10	505
1.22E-05	Human polymorphic CA dinucleotide repeat	32	5	2	27	488

Incyte gene 3458076 occurred in 7 of 522 cDNA libraries and showed strong co-expression with several of the known prostate cancer-specific genes, such as glandular kallikrein, prostate seminal protein, prostatic acid phosphatase, prostate-specific antigen, and prostate transglutaminase, as shown in Table 11. 3458076 also exhibited association with a human dinucleotide repeat flanking region, a region that flanks a polymorphic CA micro satellite repeat from the long arm of chromosome 1 (Raymond, M. H., et al. (1987) GI 2182124, GenBank). Genes in this region have not been characterized. Further,

3458076 showed coexpression with four novel Incyte genes, 1816556, 1344875, 1864683, and 1651189. These results are consistent with the notion that 3458076 is associated with prostate cancer; and 3458076 may be functionally or regulationally associated with at least four novel Incyte genes.

5

VI.   Novel Prostate Cancer-Associated Genes

Eight novel Incyte genes were identified from the data shown in Tables 5 to 12 to be associated with prostate cancer.

Nucleic acids comprising the consensus sequences of SEQ ID NOs: 1-10 of the

10   present invention were first identified from Incyte Clones 842349, 1682557, 1816556, 1864683, 2187866, 3096181, 3360806, and 3458076, respectively, and assembled according to Example III. BLAST and other motif searches were performed for SEQ ID NOs: 1-8 according to Example VII. The sequences were translated and sequence identity was sought with known sequence. Of interest, the amino acid sequence encoded by SEQ

15   ID NO: 1 from about nucleotide 195 to about nucleotide 446 showed 58% sequence identity with a subunit of a mouse RNA polymerase I, PRA16 (GI 1778684); and the amino acid sequence encoded SEQ ID NO:3 from about 185 to about 825 showed about 76% sequence identity with a Sus scrofa enamel matrix serine protease (GI 2737921). The protease activity of the enamel matrix serine protease is consistent with that of PSA and

20   kallikrein, two of the known human prostate cancer-specific genes. SEQ ID NO: 9 is an amino acid sequence coded by SEQ ID NO: 4. SEQ ID NO: 9 is 231 amino acids in length. Residue 188 to residue 209 encompass a potential transmembrane domain, and residue 1 to residue 47 is a potential signal peptide sequence. SEQ ID NO: 9 also has two potential casein kinase II phosphorylation sites at S100 and S142; one potential protein

25   kinase C phosphorylation site at S147; and a potential cell attachment sequence encompassing residues R93GD which interacts with a cell surface receptor. SEQ ID NO: 10 is an amino acid sequence coded by SEQ ID NO: 8. SEQ ID NO: 10 is 162 amino acids in length. The fragment from residue 83 to residue 99 resembles a potential BLOCK signature of Ly-6/u-PAR, a family of GPI-linked cell-surface glycoproteins. SEQ ID NO:

30   10 also has one potential N-glycosylation site at N4; one potential cAMP- and cGMP-dependent protein kinase phosphorylation site at T48; and three potential protein kinase C phosphorylation sites at T25, T34, and S44.

VII. Homology Searching for Prostate Cancer-Associated Genes and the Proteins Encoded by the Genes

Polynucleotide sequences, SEQ ID NOs: 1-8, and polypeptide sequences, SEQ ID
5   NOs: 9-10, were queried against databases derived from sources such as GenBank and SwissProt. These databases, which contain previously identified and annotated sequences, were searched for regions of similarity using Basic Local Alignment Search Tool (BLAST; Altschul, S.F. et al. (1990) J. Mol. Biol. 215: 403-410) and Smith-Waterman alignment (Smith, T. et al. (1992) Protein Engineering 5:35-51). BLAST searched for
10   matches and reported only those that satisfied the probability thresholds of 10^{-25} or less for nucleotide sequences and 10^{-8} or less for polypeptide sequences.

The polypeptide sequences were also analyzed for known motif patterns using MOTIFS, SPSCAN, BLIMPS, and Hidden Markov Model (HMM)-based protocols. MOTIFS (Genetics Computer Group, Madison, WI) searches polypeptide sequences for
15   patterns that match those defined in the Prosite Dictionary of Protein Sites and Patterns (Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221), and displays the patterns found and their corresponding literature abstracts. SPSCAN (Genetics Computer Group, Madison, WI) searches for potential signal peptide sequences using a weighted matrix method (Nielsen, H. et al. (1997) Prot. Eng. 10: 1-6). Hits with a score of 5 or greater
20   were considered. BLIMPS uses a weighted matrix analysis algorithm to search for sequence similarity between the polypeptide sequences and those contained in BLOCKS, a database consisting of short amino acid segments, or blocks, of 3-60 amino acids in length, compiled from the PROSITE database (Henikoff, S. and G. J. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Bairoch et al., supra), and those in PRINTS, a protein
25   fingerprint database based on non-redundant sequences obtained from sources such as SwissProt, GenBank, PIR, and NRL-3D (Attwood, T. K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424). For the purposes of the present invention, the BLIMPS searches reported matches with a cutoff score of 1000 or greater and a cutoff probability value of 1.0×10^{-3}. HMM-based protocols were based on a probabilistic approach and
30   searched for consensus primary structures of gene families in the protein sequences (Eddy, S.R. (1996) Cur. Opin. Str. Biol. 6:361-365; Sonnhammer, E.L.L. et al. (1997) Proteins 28:405-420). More than 500 known protein families with cutoff scores ranging from 10 to

50 bits were selected for use in this invention.

VIII. Extension of Polynucleotides

The initial primers were designed from the cDNA using OLIGO 4.06 (National

5   Biosciences, Plymouth, MN), or another appropriate program, to be about 22 to 30

nucleotides in length, to have a GC content of about 50% or more, and to anneal to the

target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides

which would result in hairpin structures and primer-primer dimerizations were avoided.

Selected human cDNA libraries (GIBCO/BRL) were used to extend the sequence. If more

10   than one extension is necessary or desired, additional sets of primers are designed to

further extend the known region.

High fidelity amplification was obtained by following the instructions for the XL-

PCR kit (Perkin Elmer) and thoroughly mixing the enzyme and reaction mix. PCR was

performed using the Peltier Thermal Cycler (PTC200; M.J. Research, Watertown, MA),

15   beginning with 40 pmol of each primer and the recommended concentrations of all other

components of the kit.

A 5 μl to 10 μl aliquot of the reaction mixture was analyzed by electrophoresis on

a low concentration (about 0.6% to 0.8%) agarose mini-gel to determine which reactions

were successful in extending the sequence. Bands thought to contain the largest products

20   were excised from the gel, purified using QIAQuick™ (QIAGEN), and trimmed of

overhangs using Klenow enzyme to facilitate religation and cloning.

After ethanol precipitation, the products were redissolved in 13 μl of ligation

buffer, 1μl T4-DNA ligase (15 units) and 1μl T4 polynucleotide kinase were added, and

the mixture was incubated at room temperature for 2 to 3 hours, or overnight at 16° C.

25   Competent E. coli cells (in 40 μl of appropriate media) were transformed with 3 μl of

ligation mixture and cultured in 80 μl of SOC medium. After incubation for one hour at

37° C, the E. coli mixture was plated on Luria Bertani (LB) agar containing 2x Carb. The

following day, several colonies were randomly picked from each plate and cultured in 150

μl of liquid LB/2x Carb medium placed in an individual well of an appropriate

30   commercially-available sterile 96-well microtiter plate.

IX. Labeling and Use of Individual Hybridization Probes

Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μCi of [γ-^{32}P] adenosine triphosphate (Amersham, Chicago, IL), and T4 polynucleotide kinase (DuPont NEN®, Boston, MA). The labeled oligonucleotides are substantially purified using a Sephadex G-25 superfine resin column (Pharmacia & Upjohn, Kalamazoo, MI). An aliquot containing 10^7 counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba 1, or Pvu II (DuPont NEN, Boston, MA).

The DNA from each digest is fractionated on a 0.7 percent agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham, NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under increasingly stringent conditions up to 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. After XOMAT AR™ film (Kodak, Rochester, NY) is exposed to the blots to film for several hours, hybridization patterns are compared visually.

What is claimed is:

1. . A method for identifying biomolecules useful in the diagnosis or treatment of a disease associated with cell proliferation, said method comprising:

5   a) examining expression patterns of polynucleotides that are expressed in a plurality of cDNA libraries, said expressed polynucleotides comprising one or more prostate cancer-specific genes and one or more genes of unknown function; and

     b) comparing the expression patterns of said prostate cancer-specific genes with the expression patterns of the genes of unknown function to identify a subset of the genes of 10 unknown function which have similar expression patterns to those of prostate cancer-specific genes.

2   The method of claim 1, wherein said polynucleotides are selected from the group consisting of expressed sequence tags (ESTs), assembled sequences, full length 15 gene coding sequences, 5' untranslated regions and 3' untranslated regions.

3.   The method of claim 1, wherein said prostate cancer-specific genes are selected from the group consisting of prostate-specific antigen, prostatic acid phosphatase, kallikrein, seminal plasma protein and prostate-specific transglutaminase.

20

4.   The method of claim 1, wherein said comparing comprises

     a) generating an occurrence data vector for each expressed polynucleotide;

     b) analyzing vectors for two or more expressed polynucleotides to determine a coexpression probability; and

25   c)determining whether the coexpression probability for said two or more expressed polynucleotides is less than a specified coexpression probability threshold.

5.   The method of claim 1, further comprising the step of translating said subset of genes of unknown function to generate corresponding polypeptides.

30

     6. A polynucleotide identified by the method of claim 1.

7. A polypeptide identified by the method of claim 5.

8. A substantially purified biomolecule for use in the diagnosis or treatment of a disease associated with cell proliferation, said biomolecule selected from the group consisting of:

(A) a polynucleotide selected from the group consisting of SEQ ID NOs: 1-8;

(B) a polynucleotide which encodes a polypeptide selected from the group consisting of SEQ ID NOs: 9-10;

(C) a polynucleotide having at least 70% identity to the polynucleotide of (A) or (B);

(D) a polynucleotide which is complementary to the polynucleotide of (A), (B), or (C);

(E) a polynucleotide comprising at least 18 sequential nucleotides of the polynucleotide of (A), (B), (C), or (D);

(F) a polypeptide selected from the group consisting of SEQ ID NOs: 9-10;

(G) a polypeptide having at least 85% identity to the polypeptide of (F); and

(H) a polypeptide comprising at least 6 sequential amino acids of the polypeptide of (F) or (G).

9. The substantially purified biomolecule of claim 8, comprising a polynucleotide sequence selected from the group consisting of:

(A) a polynucleotide selected from the group consisting of SEQ ID NOs: 1-8;

(B) a polynucleotide which encodes a polypeptide selected from the group consisting of SEQ ID NOs: 9-10;

(C) a polynucleotide having at least 70% identity to the polynucleotide of (A) or (B);

(D) a polynucleotide which is complementary to the polynucleotide of (A), (B), or (C);

(E) a polynucleotide comprising at least 18 sequential nucleotides of the polynucleotide of (A), (B), (C), or (D); and

(F) a polynucleotide which hybridizes under stringent conditions to the polynucleotide of (A), (B), (C), (D), or (E).

10. The substantially purified biomolecule of claim 8, comprising a polypeptide sequence selected from the group consisting of:

(A) a polypeptide selected from the group consisting of SEQ ID NOs:9-10;

(B) a polypeptide having at least 85% identity to the polypeptide of (A); and

5   (C) a polypeptide comprising at least 6 sequential amino acids of the polypeptide of (A) or (B).

11.   An expression vector comprising the polynucleotide of claim 9.

10   12.   A host cell comprising the expression vector of claim 11.

13.   A method for producing a polypeptide of claim 10, the method comprising the steps of:

a) culturing the host cell of claim 12 under conditions suitable for the

15   expression of the polypeptide; and

b) recovering the polypeptide from the host cell culture.

14.   A pharmaceutical composition comprising the biomolecule of claim 8 in conjunction with a suitable pharmaceutical carrier.

20

15.   An antibody which specifically binds to the polypeptide of claim 10.

16.   A method for detecting a target polynucleotide in a biological sample, the method comprising the steps of:

25   (a)   hybridizing the polynucleotide of claim 9 to the target polynucleotide to form a hybridization complex; and

(b)   detecting the hybridization complex, wherein the presence of the hybridization complex correlates with the presence of the target polynucleotide.

30   17.   A method for identifying biomolecules useful in the diagnosis or treatment of a disease, said method comprising:

a) examining expression patterns of a plurality of biomolecules that are expressed

in a plurality of cDNA libraries, said expressed biomolecules comprising one or more disease-specific biomolecules and one or more biomolecules of unknown function; and

    b) comparing the expression patterns of said disease-specific biomolecules with the expression patterns of the biomolecules of unknown function to identify a subset of the

5 biomolecules of unknown function which have similar expression patterns to those of the disease-specific biomolecules.

    18.   The method of claim 17, wherein said comparing comprises

    a) generating an occurrence data vector for each expressed biomolecule;

10   b) analyzing vectors for two or more expressed biomolecules to determine a coexpression probability; and

    c) determining whether the coexpression probability for said two or more expressed biomolecules is less than a specified coexpression probability threshold.

15   19. A polynucleotide identified by the method of claim 17.

    20. A polypeptide identified by the method of claim 17.

SEQUENCE LISTING

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<221> misc_feature
<223> Incyte clone 1816556

<400> 3
ggggccctgg acggttttct ctatccactc agtgaatttg cagaggttgg tgtagacacc  60
tggcgcgcca acttggccac acggggcttt tccgaaagac acaaggccct gcaagtaccc 120
gttgcagatc aggggccccc cagagtcacc gttgcaggag tccttctggt cttgccctcc 180
gccgggcgag gactgcagcc cgcgctcgca gccctggcag gcggcactgg tcatggaaaa 240
gggctgggcc tgcacagtct tgaggccgac caagagccag ggagccagat ggtggaggcc 300
agcctctccg tacggcaccc agagtacaac agacccttgc tcgctaacga cctcatgctc 420
```

```
atcaagttgg acgaatccgt gtccgagtct gacaccatcc ggagcatcag cattgcttcg  480
cagtgcccta ccgcggggaa ctcttgcctc gtttctggct ggggtctgct ggcgaacggc  540
agaatgccta ccgtgctgca gtgcgtgaac gtgtcggtgg tgtctgagga ggtctgcagt  600
aagctctatg acccgctgta ccacccagc atgttctgcg ccggcggagg gcaagaccag    660
aaggactcct gcaacggtga ctctggggggg cccctgatct gcaacgggta cttgcagggc  720
cttgtgtctt tcggaaaagc cccgtgtggc caagttggcg tgccaggtgt ctacaccaac  780
ctctgcaaat tcactgagtg gatagagaaa accgtccagg ccagttaact ctggggactg  840
ggaacccatg aaattgaccc ccaaatacat cctgcggaag gaattcagga atatctgttc  900
ccagcccctc gtccctnagg cccaggagtc cacnncncaa aanantcnnt ncnctaaanc  960
naggggggtac anaatccccc aanaaaggan nntnncannn angacccang ggannnnnaa 1020
aaatttccnn naaaataggc                                             1040
```

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<210> 4
<211> 2462
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte clone 1864683

<400> 4
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gaagcctatg gccagctgtc tttgtgttcc ctctcacccg cctgtcctca cagctgagac  120
tcccaggaaa ccttcagact accttcctct gccttcagca aggggcgttg cccacattct  180
ctgagggtca gtggaagaac ctagactccc attgctagag gtagaaaggg gaagggtgct  240
ggggagcagg gctggtccac agcaggtctc gtgcagcagg tacctgtggt tccgccttct  300
catctccctg agactgctcc gacccttccc tcccaggctc tgtctgatgg cccctctccc  360
tctgcaggcg ttcggatggg cagcctgggg ctgttcctgc agtgcgccat ctccctggtc  420
ttctctctgg tcatggaccg gctggtgcag cgattcggca ctcgagcagt ctatttggcc  480
agtgtggcag cttttccctgt ggctgccggt gccacatgcc tgtcccacag tgtggccgtg  540
gtgacagctt cagccgccct caccgggttc accttctcag ccctgcagat cctgccctac  600
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actggaggtg ctagcagtga ggacagcctg atgaccagct tcctgccagg ccctaagcct  720
ggagctccct tccctaatgg acacgtgggt gctggaggca gtggcctgct cccacctcca  780
cccgcgctct gcgggggcctc tgcctgtgat gtctccgtac gtgtggtggt gggtgagccc  840
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gccttcctgc tgtcccaggt ggccccatcc ctgtttatgg gctccattgt ccagctcagc  960
cagtctgtca ctgcctatat ggtgtctgcc gcagcgctgg gtctggtcgc catttacttt 1020
gctacacagg tagtatttga caagagcgac ttggccaaat actcagcgta gaaaacttcc 1080
agcacattgg ggtggagggc ctgcctcact gggtcccagc tccctgctcc tgttagcccc 1140
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cccaggctca gggttaacag ctagcctcct agttgagaca cacctagaga aggggttttttg 1440
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gtttaatgta gctcttgcat gggagtttct aggatgaaac actcctccat gggatttgaa 1560
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ctcagcccac agcactgtct ttttgctgat ccacccccct cttaccttt atcaggatgt 1680
ggcctgttgg tccttctgtt gccatcacag agacacaggc atttaaatat ttaacttatt 1740
tatttaacaa agtagaaggg aatccattgc tagcttttct gtgttggtgt ctaatatttg 1800
ggtagggtgg gggatcccca acaatcaggt cccctgagat agctggtcat tgggctgatc 1860
attgccagaa tcttcttctc ctggggtct gcccccaaa atgcctaacc caggaccttg 1920
gaaattctac tcatcccaaa tgataattcc aaatgctgtt acccaaggtt agggtgttga 1980
aggaaggtag agggtggggc ttcaggtctc aacggcttcc ctaaccaccc ctcttctctt 2040
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ggcccagcct ggttcccccc acttccactc ccctctactc tctctaggac tgggctgatg 2100
aaggcactgc ccaaaatttc ccctaccccc aactttcccc tacccccaac tttccccacc 2160
agctccacaa ccctgtttgg agctactgca ggaccagaag cacaaagtgc ggtttcccaa 2220
gcctttgtcc atctcagccc ccagagtata tctgtgcttg gggaatctca cacagaaact 2280
caggagcacc ccctgcctga gctaagggag gtcttatctc tcaggggggg tttaagtgcc 2340
gtttgcaata atgtcgtctt atttatttag cggggtgaat attttatact gtaagtgagc 2400
aatcagagta taatgtttat ggtgacaaaa ttaaaggctt tcttatatgt taaaaaaaaa 2460
aa                                                                 2462
```

<210> 5
<211> 1567
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte clone 2187866

<400> 5

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cgaaagctgg ggtggcctct gcccttctg ggttccttt tcctgcacag ctgctttctg  120
actccaccca cagctgggag caggtgccgg agccccggcc tgcatggccc tgtgaaggcc  180
actctgggcg tttgggtggg cgtgagtgcc ttcctctgct cccagcatgt ggttttctcc  240
gttggccgcc tcctggacct cctggtgcct gacatcccag agtctgtgga gatcaaagtg  300
aagcgggagt actacctggc taagcaggca ctggctgaga atgaggttct ttttggaacg  360
aacggaacaa aggatgagca gcccgagggc tcagagctca gctcccactg gacacccttc  420
acggttccca aggccagcca gctgcagcag tgacgcctgg aaggacatct ggtggtcctt  480
aggggagtgg cccctcctga gccctgcgag cagcgtcctt ttcctcttcc ctcaggcagc  540
ggctgtgtga accgctggct gctgttgtgc ctcatctctg ggcacattgc ctgcttcccc  600
ccagcgccgg cttctctcct cagagcgcct gtcactccat ccccggcagg gagggaccgt  660
cagctcacaa ggccctcttt gtttcctgct cccagacata agcccaaggg gcccctgcac  720
ccaagggacc ctgtccctcg gtggcctccc caggccctg gacacgacag ttctcctcag  780
gcaggtgggc tttgtggtcc tcgccgcccc tggccacatc gccctctcct cttacacctg  840
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gccccttcct gccctgtttg cgcagggaca tcacccacat gccccagctc tcggaccctg  960
cagctctgtg tcccaggcca cagcaaaggt ctgttgaacc cctccctcca ttcccagtta 1020
tctgggtcct ctggattctt ctgtttcttg aatcaggctc tgctttcccc ctagccacta 1080
caggcagcct ctgacagtgc cgctttactt gcattctgca gcaattacat gtgtcctttt 1140
gatccttgcc caacttccct ccctctccca gctcctggcc cctggcccag ggccctctt 1200
gctgttttta cctctgttcc ttggggccta gtacccagca agcacccaaa tgggggaggt 1260
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tgtggctgtc tgcttgctaa aaaatggatc tgtgctcatc tgtattgatg tccttggagt 1500
tctacaagtg gaacttaagt gtcaaaaaga atatgtggtt tttaggggga tcctctagag 1560
tcgacct                                                           1567
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<210> 6
<211> 1354
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte clone 3096181

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<400> 6
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agataagttc cgtaagacaa gagatactct cagaaaaaag tcatcggctt tagaaactct 120
ccaaacgacc taagccaaac acagcagcaa ataaaggaaa tgaaagagat gtatgaaat  180
gcagaagata aagtgaataa ttccactgga aagtggagct gtgtagaaga gaggatatgt 240
catctccaac atgaaaatcc gtgccttgaa cagcaactag atgatgttca tcagaaagag 300
gatcataaag agatagtaac taatatccaa agaggcttta ttgagagtag aaagaaagac 360
ctcatgctag aagagaaaag taagaagcta atgaatgaat gtgatcattt aaaagaaagt 420
ctctttcaat atgagagaga gaaagcagaa agagtagtgg ttgtgagaca acttcaacaa 480
gaagcggctg acagccctaa aaaaattaac tatgttagag tctccactgg aaggtatatc 540
acgttgtcat attaatttgg ttgagacaca ggtcccaaag aagaaattat ttcaagtgga 600
aagtcaattt gatgatctta tggcggagaa ggaagctgta tcttcaaaat gtgtcaattt 660
ggctaaagag aatcaagttt ttcaacagga gttattatct atgaaaaaag cacaacagga 720
atgtgaaaaa tttgaggagg ataaaagatg ttggaagaag aaatattaaa tcttaagaca 780
catatggaaa acagtatggt agaacttagt aaactacaag aatataaatc agagctagat 840
gaaaaggcaa tgcaggcagt agaaaaatta gaagaaatcc atttacagga acaagcacaa 900
tataaaaaac aattagagca gttaaacaag gatataatac agcttcacta aataagaagg 960
aactcacact taaagatgtg gaatgtaaat tctacaaaat gaaaactgct tatgaagagg 1020
ttacaactga gttagaagaa tataaggaag cctttgcagc agcattgana gctaacagtt 1080
ccatgtctaa nanattaact aaatcgaata aganaatagc aatgatcagt atcagctctt 1140
tatggngana gagcaggtga natattttct cagcactctt cctacnnggc gaggtcgaga 1200
gtcaccttgt gttgantatn cctacttngt ataggngctc agtcaganga tataatnctt 1260
ccaaatngcc cgtntggggt tcctancttt caggngcctn cagagnttcc ataattnaac 1320
ngnccnaggn gtanctttga nctgagggcc nntt                            1354
```

```
<210> 7
<211> 2426
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte clone 3360806

<400> 7
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aaganccccc gccagcttcc cacacctcat acgcagccac atctgcccta ttctccatgc 120
tttccagctt gcctgcccctt cctcatctct ccctgcctgt gcagacctcc acccttcttt 180
cctccacccc tccatccccc aatgcttgta gaccttccat tcattccgtc tcatcgtgcg 240
tggtctctga tcgtccatca cctgaccttc tccaggactg tcttctcacc cttccccact 300
ccctggtccc cgggagcagc tccttctgcc cgactcactc acagtgcagg gaaaggaggc 360
agggaaaaga ccaggattct gtgagttctg aggttgccac acacaaagaa gctgtggttt 420
ctctgcctcg gccactgatg agactaaaac tggcttcccc ttggagacgg cagatttcag 480
gctgatccct gcttaagccc tctcatcccc acgctggtcc tggtattgat acaagaccca 540
gctggtgaca aagcctccaa tcctggggt ccacgagcct gggcctgaca ttcccagaac 600
taccgccagt ggcgccaggc ccccacagtc tgtggccgtg gtcttagccc ccagttccac 660
tctggatggg cctgtgacac cccaaagaga agaaggggac tctggatagg gtccccacat 720
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gagctgagca ttctgcttgc tgcagtagac gggtcgcctt ttgcccatac cgaaattttc 840
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cgttccttgt aaatccaaat gtttctatat tgtagctttg cttaaaatgg gggtcggccc 1020
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cgaccccgag gcctcgtgga cgtgggagag agtgtggtgg gaagtcttga gcggaggagg 1140
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cgcctgcccc gcggaagacc gcccagtgag ccagcccca ccttccaggc gccttcgccc 1260
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tggggatcca accaacttgt atcgagtggg cggggcaacg gctccccatt tttcccgagc 1320
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cccaaccaga ccccacccag gcacattagc gaccaggctt gggcttcccc agcgccccac 1440
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ccaccatccc aggactttgg gcaagtcacc cgcactccct gggcctcggt ttccccatct 1680
gtaaaatgat ggtaataata cttcacctac ctcatagggg aggttgtgag gccaccatca 1740
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gctccccgga gccccagggg tggggattgg tggccactgt ttgtatgttc ttgagtgcaa 2160
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ttccaccaca catacccctct aagccagcaa gatttcctct ttgcaaaatc atatttttgt 2340
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<210> 8
<211> 510
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte clone 3458076

<400> 8
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gtccccactg aggtgcccca cagcagcagg tgttgagcat gggctgagaa gctggaccgg 180
caccaaaggg ctggcagaaa tgggcgcctg gctgattcct aggcagttgg cggcagcaag 240
gaggagaggc cgcagcttct ggagcagagc cgagacgaag cagttctgga gtgcctgaac 300
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gctgcggcac cggaaagccc agctcttgct ggtcaacctg ctaacctttg gcctggaggt 420
gtgtttggcc gcagattcac ctatgtgccg cctctgtgct ggaatggggg tagaggagaa 480
gttcatgacc atggtgctgg gatttggtcc                                  510
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<210> 9
<211> 231
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte cone 1864683

<400> 9
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Met Gly Ser Leu Gly Leu Phe Leu Gln Cys Ala Ile Ser Leu Val
 1               5                  10                  15
Phe Ser Leu Val Met Asp Arg Leu Val Gln Arg Phe Gly Thr Arg
                20                  25                  30
```

```
Ala Val Tyr Leu Ala Ser Val Ala Ala Phe Pro Val Ala Ala Gly
             35                  40                  45
Ala Thr Cys Leu Ser His Ser Val Ala Val Val Thr Ala Ser Ala
             50                  55                  60
Ala Leu Thr Gly Phe Thr Phe Ser Ala Leu Gln Ile Leu Pro Tyr
             65                  70                  75
Thr Leu Ala Ser Leu Tyr His Arg Glu Lys Gln Val Phe Leu Pro
             80                  85                  90
Lys Tyr Arg Gly Asp Thr Gly Gly Ala Ser Ser Glu Asp Ser Leu
             95                 100                 105
Met Thr Ser Phe Leu Pro Gly Pro Lys Pro Gly Ala Pro Phe Pro
            110                 115                 120
Asn Gly His Val Gly Ala Gly Gly Ser Gly Leu Leu Pro Pro Pro
            125                 130                 135
Pro Ala Leu Cys Gly Ala Ser Ala Cys Asp Val Ser Val Arg Val
            140                 145                 150
Val Val Gly Glu Pro Thr Glu Ala Arg Val Val Pro Gly Arg Gly
            155                 160                 165
Ile Cys Leu Asp Leu Ala Ile Leu Asp Ser Ala Phe Leu Leu Ser
            170                 175                 180
Gln Val Ala Pro Ser Leu Phe Met Gly Ser Ile Val Gln Leu Ser
            185                 190                 195
Gln Ser Val Thr Ala Tyr Met Val Ser Ala Ala Ala Leu Gly Leu
            200                 205                 210
Val Ala Ile Tyr Phe Ala Thr Gln Val Val Phe Asp Lys Ser Asp
            215                 220                 225
Leu Ala Lys Tyr Ser Ala
            230
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<210> 10
<211> 162
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte clone 3458076

<400> 10
Met Val Met Asn Phe Ser Ser Thr Pro Ile Pro Ala Gln Arg Arg
 1               5                  10                  15
His Ile Gly Glu Ser Ala Ala Lys His Thr Ser Arg Pro Lys Val
             20                  25                  30
Ser Arg Leu Thr Ser Lys Ser Trp Ala Phe Arg Cys Arg Ser Arg
             35                  40                  45
Arg Leu Thr His Ser Leu Trp Thr Ile Val Gly Gln Ala Gly Arg
             50                  55                  60
Ala Gln Gly Ala Val Gln Ala Leu Gln Asn Cys Phe Val Ser Ala
             65                  70                  75
Leu Leu Gln Lys Leu Arg Pro Leu Leu Leu Ala Ala Ala Asn Cys
             80                  85                  90
Leu Gly Ile Ser Gln Ala Pro Ile Ser Ala Ser Pro Leu Val Pro
             95                 100                 105
Val Gln Leu Leu Ser Pro Cys Ser Thr Pro Ala Ala Val Gly His
```

```
          110                     115                     120
Leu Ser Gly Asp Thr Ser His His Ser Asp Pro Gly Arg Gly Ala
          125                     130                     135
Arg Leu Ser Pro Gly Ala Ser Ala Xaa Arg Leu Val Pro Pro Pro
          140                     145                     150
Leu Pro Cys Pro Pro Arg Ala Pro Ala Gly Gly Phe
          155                     160
```